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# Modelling the impacts of the foliar pathogen, *Phaeocryptopus gaeumannii*, on Douglas-fir physiology: net canopy carbon assimilation, needle abscission and growth

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## Abstract

This paper describes the parameterisation, testing and implementation of needle-level stomatal conductance ( $g_s$ ) and net CO<sub>2</sub> assimilation ( $A_{net}$ ) models that include the physiological impacts of the Douglas-fir pathogen, *Phaeocryptopus gaeumannii*. Hourly estimates of  $g_s$  were modelled by assuming that stomata regulate water flux such that plant water potential is maintained above a critical threshold, and  $A_{net}$  was modelled based on the kinetics of photochemistry. The model was tested using summer field measurements from trees at three western Oregon Douglas-fir (*Pseudotsuga menziesii*) plantations with varying levels of *P. gaeumannii*, and showed a high degree of accuracy:  $r^2 = 0.777$  and  $0.792$  for  $g_s$  and  $A_{net}$ , respectively. Instantaneous needle-level estimates of  $g_s$  and  $A_{net}$  were also scaled-up to a whole-canopy estimate for a 10-month period (July 1998–April 1999). At all three sites, a significant seasonality in  $A_{net}$  was observed, with the highest rates occurring during the summer months (up to  $400 \text{ g CO}_2 \text{ m}^{-2} \text{ LA}$ ) declining to near or below zero during the winter. The presence of *P. gaeumannii* had a significant impact on needle- and whole-canopy  $A_{net}$ , and for the needle age classes where colonisation levels reached 25% pseudothecia density (i.e. percent of stomata with visible fruiting bodies), estimated total carbon budgets were negative. However, at the whole-canopy level all trees maintained a positive carbon budget due to the large contribution from current year needles that remain unaffected by the fungus for the first 6 months of development, or until the emergence of pseudothecia. Furthermore, the abscission of the older, more-heavily diseased foliage, shortly after becoming a carbon sink, has a significant mitigating effect on whole-canopy  $A_{net}$ . For example, at the high-disease site, *P. gaeumannii*-associated reductions in  $A_{net}$  per unit leaf were estimated to reduce whole-canopy  $A_{net}$  by ca. 110% without needle abscission, but this was reduced to 85% when older, more-heavily diseased needles were abscised.

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## 1. Introduction

Douglas-fir (*Pseudotsuga menziesii*) forest plantations are some of the most productive in the world (McMurtrie, 1993), with over 15 million ha currently

in production (Hermann and Lavender, 1999). However, over the past 10 years, Swiss needle cast disease (SNC), caused by the parasitic foliar fungus, *Phaeocryptopus gaeumannii*, has intensified in forest plantations of the Pacific Northwest of the United States (PNW). The area affected by SNC increased from 52,600 to 119,500 ha between 1996 and 1999 (Hansen et al., 2000), potentially limiting the productivity of these stands. Indeed, volume growth has recently been reported to be 23–50% less than expected in these PNW forest plantations (Maguire et al., 2002); however, it is unknown if SNC is responsible for the observed growth losses. Historically, *P. gaeumannii* has been considered a weak pathogen with limited impact on tree growth (Boyce, 1940; Hood, 1982); however, given the current accelerated rates of fungal development i.e. fruiting occurring on current year (Hansen et al., 2000) rather than 3- or 4-year-old needles (Boyce, 1940; Hood, 1982) and recent elucidation of the mechanism of *P. gaeumannii*'s impact on Douglas-fir physiology (Manter et al., 2000), we chose to quantify the potential influence of *P. gaeumannii* on whole-tree physiology and growth using a modelling approach.

The objective of this study was to parameterize and test a model of Douglas-fir net CO<sub>2</sub> assimilation with varying levels of *P. gaeumannii* colonisation. Once validated, this model could then be used to determine: (i) the impact of *P. gaeumannii* on Douglas-fir physiology at larger time and spatial scales, (ii) if *P. gaeumannii*'s impact is correlated with the current growth losses in coastal Oregon Douglas-fir plantations—testing the hypothesis that SNC is responsible for the observed growth losses, and (iii) the impact as disease level increases or decreases.

## 2. Model development and testing

### 2.1. Plant material

The plant material necessary for estimation of model parameters consisted of Douglas-fir seedlings and field trees from two separate studies. The field trees consisted of 12–15-year-old trees from three western Oregon plantations. Two of the plantations are located in the Coast Range (i.e. Beaver and Hebo), and these sites have severe and moderate levels of *P.*

*gaeumannii* infection, respectively. The third plantation is from the Willamette Valley with low levels of *P. gaeumannii* infection (i.e. Mac). At each site, paired permanent plots were created on north- and south-facing slopes (10–30%), and within each plot six infected (i.e. no spray) trees and six uninfected (i.e. spray) control trees were measured. Control trees were kept disease-free by spraying foliage with chlorothalonil (Bravo 720, rate = 66 ml/3.78 l, applied until run-off) by means of a backpack sprayer. Fungicide applications were conducted in 1998 and 1999 at bud break (90% trees had broken bud) and 1 month following bud break.

Two-year-old Douglas-fir seedlings (open-pollinated Oregon north-coast seed source, Starker Forests Inc., Corvallis, OR, USA) with different levels of *P. gaeumannii* infection were also utilised. To vary rates of infection, seedlings were incubated for 2, 4, or 8 weeks in the understory of *P. gaeumannii*-infected trees (Salal plot, see Hansen et al., 2000) from May to July 1999. Except during the inoculation period, all seedlings were stored in a cold frame at the Oregon State University Botany Farm (Corvallis, OR, USA), fertilised with Osmocote Pro 18-8-8 (Scotts-Sierra Horticultural Products Co., Marysville, OH, USA) and irrigated as needed.

### 2.2. Model parameterisation

Estimates of photobiochemical parameters (e.g.  $V_{\text{Cmax}}$ , maximum rate of carboxylation;  $J_{\text{max}}$ , maximum rate of electron transport; and  $R_{\text{d}}$ , respiration in the presence of light) were calculated from  $A/C_i$  curves (i.e. net CO<sub>2</sub> assimilation measured over a range of calculated internal CO<sub>2</sub> concentrations or  $C_i$ ) according to the model proposed by Farquhar et al. (1980). A LiCor 6400 gas exchange system (LiCor, Inc., Lincoln, NE, USA) was used for these measurements. During measurements, cuvette conditions were maintained at photosynthetic photon flux density (400–700 nm, PPF) = 2000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ , temperature = 25 °C, [H<sub>2</sub>O vapor]  $\geq$  14 mmol mol air<sup>-1</sup>, and flow rate = 100 mmol m<sup>-2</sup> s<sup>-1</sup>, unless otherwise noted.  $A/C_i$  response curves were measured by varying the cuvette CO<sub>2</sub> concentration, allowing equilibration to a steady state ([CO<sub>2</sub>] coefficient of variation < 2%), and logging measurements every 10 s for 1 min. CO<sub>2</sub> was varied in the following order: 40, 30, 20, 60, 80, 100,

120, 160, and 200 Pa. Temperature dependencies photochemical parameters were determined from  $A/C_i$  curves conducted at air temperatures of 10, 18, 23, 25, 28, 30, and 35 °C following a 45 min equilibration period.

Infection levels of *P. gaeumannii* were estimated by the presence of pseudothecia (i.e. fruiting bodies) emerging from stomata. Values are reported as the percent of stomata occupied with pseudothecia (i.e. pseudothecia density or PD), calculated as the mean value of for each sampled tree based on the sampling procedures outlined in Manter et al. (2000).

Following  $A/C_i$  analysis of the field trees (12 per site) in June 1998, 2 g FW of 1-year-old needles adjacent to the cuvette were collected and used to determine total foliar *N* content (% w/w). All analyses were performed by the Central Analytical Laboratory (Crop and Soil Science Department, Oregon State University, Corvallis, OR, USA) using a Leco CNS 2000.

### 2.3. Model derivation

The needle-level gas exchange model consists of two separate but related processes. First,  $g_s$  is calculated assuming that stomatal regulation operates in such a manner as to maintain plant water potential above a threshold level (Bond and Kavanagh, 1999). Second,  $A_{\text{net}}$  is estimated based on the model proposed by Farquhar et al. (1980).

Several authors (e.g. Jones and Sutherland, 1991; Dewar, 1995; Whitehead, 1998; Bond and Kavanagh, 1999) have suggested that stomatal behavior is dependent on the water supply capacity of the vascular system, and can be described mathematically:

$$g_s \times \text{VPD} = K_L(\Psi_{\text{soil}} - \Psi_{\text{leaf}}) \quad (1)$$

where  $g_s$  is the stomatal conductance ( $\text{mmol m}^{-2} \text{s}^{-1}$ ), VPD the leaf-to-air vapor pressure gradient (Pa),  $K_L$  the leaf-specific hydraulic conductance ( $\text{mmol m}^{-2} \text{s}^{-1} \text{MPa}^{-1}$ ), and  $\Psi_{\text{soil}}$  and  $\Psi_{\text{leaf}}$  are the water potential of the soil and leaf (MPa), respectively. Estimation of  $g_s$  using Eq. (1) was similar to that proposed by Bond and Kavanagh (1999). Calculations were as follows:

Step 1. A maximum stomatal conductance ( $g_{s_{\text{max}}}$ ) based on the presence of *P. gaeumannii* was calculated using Eq. (2) (adapted from Manter

et al., 2000):

$$g_{s_{\text{max}}} = 150 \left( \frac{100 - (0.696 + 1.720 \times \text{PD})}{100} \right) \quad (2)$$

where PD is the percent of stomata occupied with *P. gaeumannii* pseudothecia and 150 is a typical  $g_{s_{\text{max}}}$  for Douglas-fir (Bond and Kavanagh, 1999).

Step 2. A preliminary value of  $\Psi_{\text{leaf}}$  was estimated using a reorganised Eq. (1):

$$\Psi_{\text{leaf}} = \Psi_{\text{soil}} - \left( \frac{g_{s_{\text{max}}} \times \text{VPD}}{K_L} \right) \quad (3)$$

Based on the results of water flux studies in Douglas-fir branches (D.K. Manter and K.L. Kavanagh, unpublished data), a *P. gaeumannii*-adjusted whole-tree  $K_L$  was calculated as:

$$K_L = (111.9 - 0.976 \times \text{LW}) \times 0.001 \quad (4)$$

where LW is the percent of latewood in each sapwood annual ring, and 0.001 is scaling factor relating branch and whole-tree  $K_L$ .

Step 3. If  $\Psi_{\text{leaf}} < \Psi_{\text{threshold}}$

Step 3a.  $\Psi_{\text{leaf}} = \Psi_{\text{threshold}}$

Step 3b.  $g_s = K_L(\Psi_{\text{soil}} - \Psi_{\text{leaf}})/\text{VPD}$

Step 4. If  $\Psi_{\text{leaf}} > \Psi_{\text{threshold}}$

Step 4a.  $g_s = g_{s_{\text{max}}}$

Step 5. A light-limited  $g_s$  ( $g_{s_{\text{ppfd}}}$ ), based on data from Leverenz (1981), was calculated:

$$g_{s_{\text{ppfd}}} = g_{s_{\text{max}}}(1 - \exp(-0.017 \times \text{PPFD})) \quad (5)$$

Step 6. Finally, the modelled stomatal conductance ( $g_{s_{\text{model}}}$ ) was set equal to

$$g_{s_{\text{model}}} = \min(g_s, g_{s_{\text{ppfd}}}) \quad (6)$$

The model of photosynthesis is based on the biochemistry model outlined in Farquhar et al. (1980), with some modifications due to the impacts of *P. gaeumannii* infection. As suggested by Farquhar et al. (1980), net  $\text{CO}_2$  assimilation can be expressed mathematically as:

$$A = V_c - 0.5 \times V_o - R_d = V_c \left( 1 - \frac{0.5 \times O}{\tau C_i} \right) - R_d \quad (7)$$

where  $V_c$  and  $V_o$  are rubisco carboxylation and oxygenation rates ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ), respectively.  $C_i$  and  $O$  are the intercellular partial pressures (Pa) of  $\text{CO}_2$  and  $\text{O}_2$ ,  $R_d$  the respiration rate ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) in the presence of light, and  $\tau$  ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) is the specificity factor for rubisco (Jordan and Ogren, 1984). The rate of  $V_c$  is limited by either the rubisco activity ( $W_c$ ) or electron transport ( $W_j$ ), therefore Eq. (7) becomes:

$$A = \left(1 - \frac{0.5 \times O}{\tau C_i}\right) \min(W_c, W_j) - R_d \quad (8)$$

$$W_c = \left(\frac{V_{c\max} C_i}{C_i + K_c(1 + O/K_o)}\right) \quad (9)$$

$$W_j = \left(\frac{J C_i}{4(C_i + O/\tau)}\right) \quad (10)$$

$$J = \frac{\alpha I}{(1 + (\alpha^2 I^2 / J_{\max}^2))^{0.5}} \quad (11)$$

where  $V_{c\max}$  is the maximum rate of carboxylation ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ),  $K_c$  and  $K_o$  the Michaelis–Menton constants for carboxylation and oxygenation, respectively,  $J$  the rate of electron transport ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ),  $\alpha$  the efficiency of light energy conversion on an incident light basis ( $\text{mole}^{-1}$  absorbed photons) and  $J_{\max}$  is the maximum rate of electron transport ( $\mu\text{mol m}^{-2} \text{s}^{-1}$ ).

Before proceeding with the photosynthesis estimation procedure several unknown parameters must first be defined (i.e.  $K_c$ ,  $K_o$ ,  $t$ ,  $R_d$ ,  $V_{c\max}$ , and  $J_{\max}$ ). As in Harley et al. (1992) we chose to model  $K_c$ ,  $K_o$  and  $\tau$  based on the temperature dependencies outlined in Jordan and Ogren (1984) using Eq. (12):

$$\text{Parameter} = \exp\left(\frac{c - \Delta H_a}{RT_k}\right) \quad (12)$$

where  $c$  is a scaling constant,  $\Delta H_a$  an activation energy,  $R$  the universal gas constant and  $T_k$  is the leaf temperature (K) (see Table 1, Harley et al., 1992). Whereas the temperature dependencies of  $V_{c\max}$  and  $J_{\max}$  can be mathematically expressed as

$$\text{Parameter} = \frac{\exp(c - \Delta H_a/RT_k)}{1 + \exp((\Delta S T_k - \Delta H_d)/RT_k)} \quad (13)$$

where  $\Delta H_d$  is the energy of deactivation and  $\Delta S$  is an entropy term. All parameter estimates used in Eq. (13) are shown in Fig. 1.

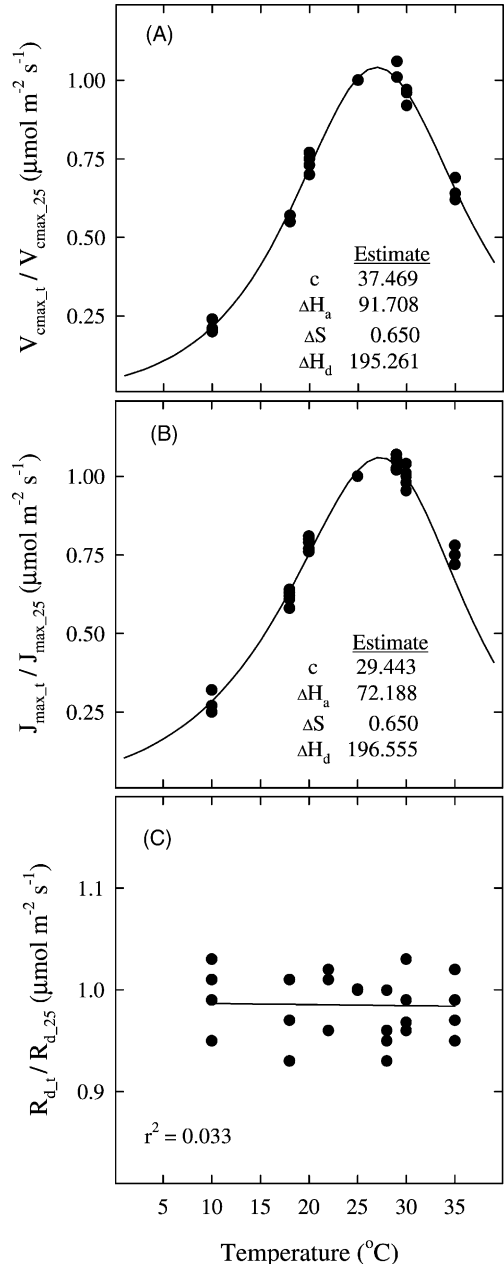


Fig. 1. Temperature dependency of the maximum rates of carboxylation ( $V_{c\max}$ ) and electron transport ( $J_{\max}$ ), and respiration rate in the presence of light ( $R_d$ ) for six 2-year-old Douglas-fir seedlings infected with varying amounts of *P. gaeumannii* (i.e. PD was ca. 0, 5, 10, 20, 40 and 50%). Parameters were estimated from  $A/C_i$  curves at each air temperature following a 45 min equilibration time using a LiCor 6400. To account for fungal-induced changes, all parameters were normalised by dividing by parameter estimates at 25°C. Measurements were conducted in April 2000.

Table 1

Observed maximum net CO<sub>2</sub> assimilation ( $A_{net}$ ,  $\mu\text{mol m}^{-2} \text{s}^{-1}$ ) and PD (%) from three western Oregon Douglas-fir plantations

Age class	Mac sites		Hebo sites		Beaver sites	
	$A_{net}$	PD	$A_{net}$	PD	$A_{net}$	PD
Current year	11.0 (1.5) a	0.0 (0.0) x	10.3 (0.9) a	1.5 (0.5) x	8.5 (1.1) a	2.2 (0.6) x
1 year	9.2 (1.3) a	3.1 (2.3) y	7.6 (1.2) b	12.0 (1.4) y	4.9 (0.7) b	24.7 (5.1) y
2 year	10.3 (1.6) a	0.2 (0.1) x	3.8 (0.8) c	18.6 (2.1) z	2.1 (0.4) c	45.0 (6.1) z

Using the four diurnal measurements of gas exchange for each site (June, July, August, and September 1998) an average of maximum  $A_{net}$  and PD (pseudothecia density) was calculated for each day from the six un-sprayed trees (north and south plots combined) at each site. Standard errors are in parentheses. For each parameter, means for the 1- and 2-year-old age classes were compared with the current year age class using a paired *t*-test. Means within a column with different letters are significantly different  $P \leq 0.05$ .

For  $V_{cmax}$ ,  $J_{max}$ , and  $R_d$ , a Douglas-fir specific temperature dependency was estimated using the 2-year-old seedlings (Fig. 1). Due to *P. gaeumannii* impacts on  $V_{cmax}$  and  $J_{max}$  (see Eqs. (14) and (15)), all values for a given seedling (Fig. 1) were normalised by parameter estimates at 25 °C (i.e.  $V_{cmax\_25}$ ,  $J_{max\_25}$ , and  $R_{d\_25}$ ). In previous studies (e.g. Farquhar et al., 1980; Harley et al., 1992),  $V_{cmax}$  has been estimated from leaf nitrogen concentrations. Based on an assessment of this relationship in 1-year-old needles from 36 coastal Oregon Douglas-fir trees (12 from each of the high-, medium- and low-disease plantations), we found no relationship between leaf nitrogen concentration and  $V_{cmax}$  (Fig. 2). However, a

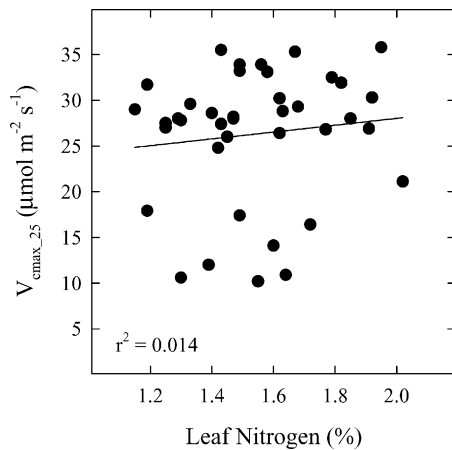


Fig. 2. Relationship between leaf nitrogen concentration and the maximum rate of carboxylation at 25 °C ( $V_{cmax\_25}$ ) in 1-year-old needles from three coastal Oregon Douglas-fir plantations. Six sprayed and six un-sprayed (PD ranged from 0 to 30%) trees were sampled from western Oregon Douglas-fir plantations ( $n = 36$ ). Measurements were conducted in June 1998.

strong correlation between  $V_{cmax}$  and *P. gaeumannii* infection levels was observed (Fig. 3A); therefore,  $V_{cmax\_25}$  was estimated using Eq. (14):

$$V_{cmax\_25} = 30.641 \times \exp(-0.041 \times \text{PD}) \quad (14)$$

Previous studies (e.g. Harley et al., 1992; Wullschlegel, 1993) report a strong correlation between  $V_{cmax\_25}$  and  $J_{max\_25}$ . In our sample population, a significant relationship was also observed (Fig. 3B), and is defined by Eq. (15):

$$J_{max\_25} = 23.914 + 1.955 \times V_{cmax\_25} \quad (15)$$

Finally, because neither temperature (Fig. 1C, also see Harley et al., 1992) nor *P. gaeumannii* infection level (Fig. 3C) had a significant effect on  $R_d$ ,  $R_d$  was held constant at  $1.64 \text{ mmol m}^{-2} \text{ s}^{-1}$ .

All climatic data required for model parameterisation (RH, *T*, and PPF) were collected using HOBO dataloggers Spectrum Technologies, Inc. (Plainfield, IL, USA). One set of dataloggers was installed at a height of 1.5 m in an opening (radius > 5 m) at each study site. Due to the wide tree spacing, open canopies, and low needle retention in these stands, within-canopy changes in light penetration were negligible and not incorporated into the model. The remaining unknown, and driving variable for the above biochemical equations, is  $C_i$ , which results from the interaction of  $A_{net}$  and  $g_s$  based on the flux equation:

$$C_i = C_a - \frac{A}{g_s} \quad (16)$$

where  $C_a$  is the atmospheric CO<sub>2</sub> concentration (assumed to be 35.5 Pa) outside the leaf boundary layer. Due to the interactive effects of  $A_{net}$  and  $g_s$  on  $C_i$ , we cannot estimate  $C_i$  independent of  $A_{net}$  and  $g_s$ .

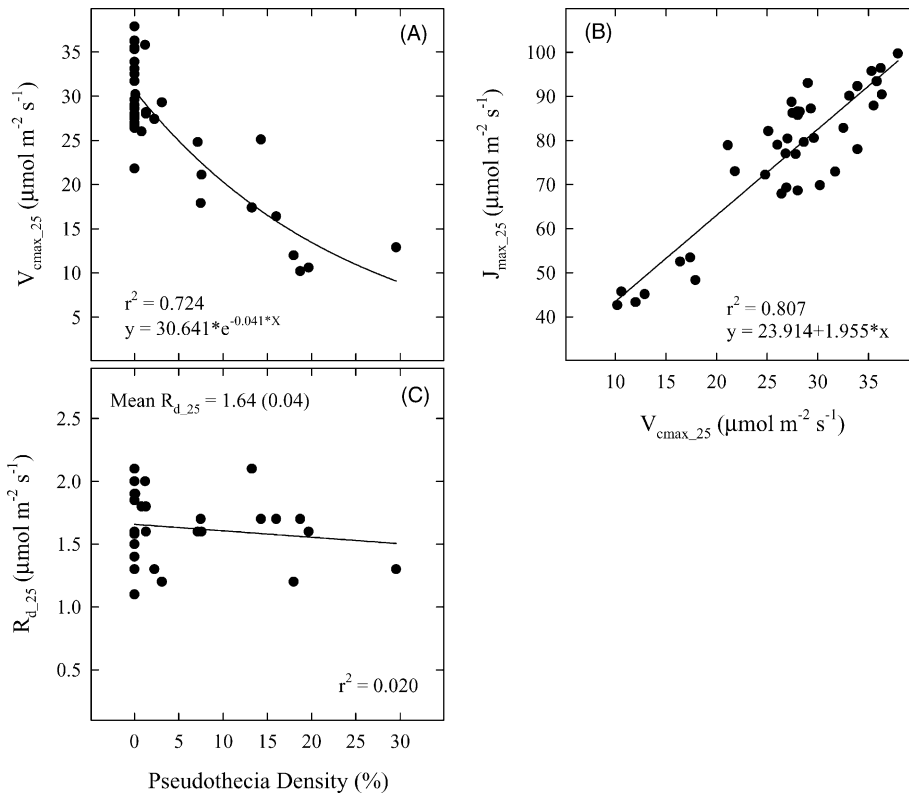


Fig. 3. Relationships used to estimate Farquhar-biochemistry parameters, i.e. the maximum rates of carboxylation ( $V_{cmax}$ ) and electron transport ( $J_{max}$ ), and respiration rate in the presence of light ( $R_d$ ), in 1-year-old needles from three coastal Oregon Douglas-fir plantations. Six sprayed and six un-sprayed (PD) ranged from 0 to 30% trees were sampled from western Oregon Douglas-fir plantations ( $n = 36$ ). Measurements were conducted in June 1998.

This problem was overcome by first estimating  $g_s$  (see stomatal conductance sub-model), then simultaneously solving for the two unknowns ( $A_{net}$  and  $C_i$ ) using Eqs. (8) and (16). For all observations, unique values of  $A_{net}$  and  $C_i$  were estimated using the model procedure in the SAS statistical package (SAS Institute Inc., Cary, NC, USA).

#### 2.4. Model validation

Model estimates were tested against diurnal gas exchange measurements from at least three diseased and three healthy trees (i.e. sprayed with the fungicide chlorothalonil) at each site. On each sample date, gas exchange was measured on the three youngest needle age classes from one sun-exposed, lower-canopy branch. 1998 measurements of  $\text{CO}_2$  and water fluxes were recorded using a LiCor 6200 portable infrared

gas exchange analyser, and 1999 measurements consisted of water flux using a LiCor 1600.

Fig. 4 shows a plot of the predicted versus measured stomatal conductance values. Overall, modelled values were highly correlated with observed values ( $r^2 = 0.777$ ); however, the model tended to slightly underestimate  $g_s$  (slope = 0.818). Regression analysis showed that none of the treatments (i.e. site, *P. gaeumannii* infection level, needle age) significantly affected the relationship between observed and predicted  $g_s$  values (data not shown). The major limitation of using our model to predict  $g_s$  arises from an overestimation of  $g_s$  during the mid-morning hours. For example, Fig. 5 depicts a typical diurnal curve for both observed and predicted  $g_s$  at a low-disease site (Mac-north). Clearly, observed values of stomatal conductance decline earlier than the modelled values. Two factors may be responsible. First, our estimates



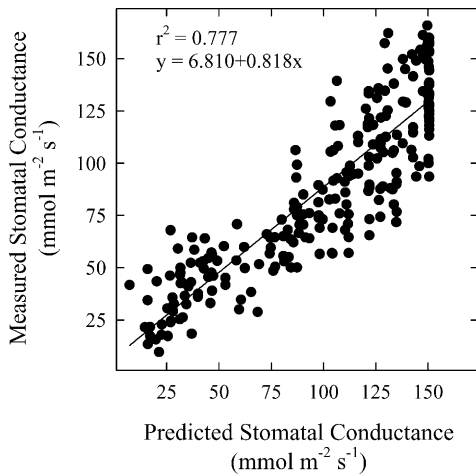


Fig. 4. Measured vs. predicted stomatal conductance ( $g_s$ ) for current year, 1- and 2-year-old needles with varying levels of *P. gaemannii* infection (PD) ranged from 0 to 61% from three western Oregon Douglas-fir plantations. Each point is the mean of six trees ( $n = 289$ ) measured in 1998 and 1999.

of  $\Psi_{\text{leaf}}$  are based on instantaneous estimates of water loss, which may overestimate  $\Psi_{\text{leaf}}$  due to the loss of water prior to the current time-step. Second, our model assumes that stomatal closure occurs only at the critical value of  $\Psi_{\text{leaf}}$  ( $-2.1$  MPa); however,  $g_s$  may begin to decline at higher  $\Psi_{\text{leaf}}$  (Webb, 1991).

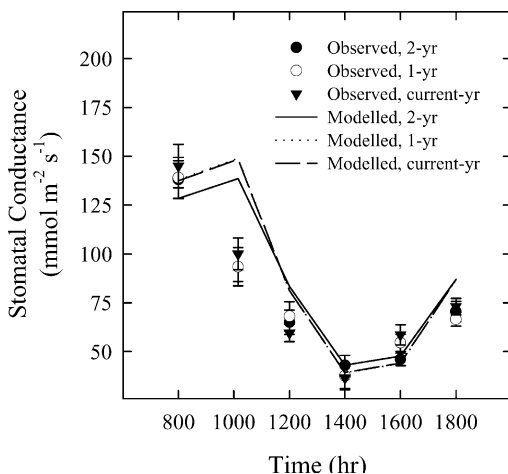


Fig. 5. Diurnal pattern of measured and predicted stomatal conductance ( $g_s$ ) for sprayed current year, 1- and 2-year-old needles (mean PD was 0, 0 and 4%, respectively) from the low-disease site (Mac-north) on 9 July 1998. Each observation is the mean and standard error for three trees.

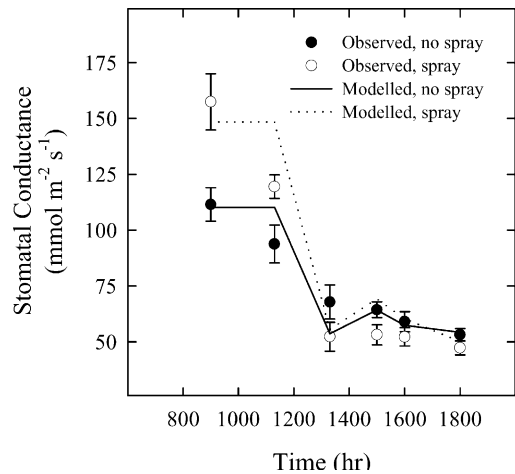


Fig. 6. Diurnal pattern of measured and predicted stomatal conductance ( $g_s$ ) for sprayed (mean PD was 0%) and un-sprayed (mean PD was 15%) 1-year-old needles from the high-disease site (Beaver-south) on 7 July 1998. Each observation is the mean and standard error for three trees.

Some models (e.g. Williams et al., 1996) have overcome the former limitation by including an algorithm for determining  $\Psi_{\text{leaf}}$  based on previous amounts of water loss and cell elasticity. However, for the purpose of modelling the relative impacts of *P. gaemannii* on needle physiology this is not necessary. Overall, our model was well correlated with observed values of stomatal conductance, and shows the impact of *P. gaemannii* infection on  $g_s$ . For example, Fig. 6 compares the diurnal patterns of  $g_s$  on a typical day from a healthy and diseased tree at a high-disease site (Beaver-south). Also evident in Fig. 6 is the limitation imposed on maximum (i.e. early morning)  $g_s$  by *P. gaemannii* infection.

Similar to  $g_s$ , our estimates of  $A_{\text{net}}$  were significantly correlated with observed values ( $r^2 = 0.792$ ), again with a slight underestimation (slope = 0.820) (Fig. 7). The observed time-lag and over-estimation of early morning values were not as apparent for  $A_{\text{net}}$  estimates (Fig. 8) as compared to those for  $g_s$ . Fig. 8 shows a typical diurnal curve for three age classes of needles with no *P. gaemannii* infection from a low-disease site (Mac-north). For this, and all days measured, a significant difference in maximum  $A_{\text{net}}$  between needle age classes was detected for needles from the high- (Beaver) and moderate-disease (Hebo) sites but not the low-disease site (Mac) (Table 1). The association

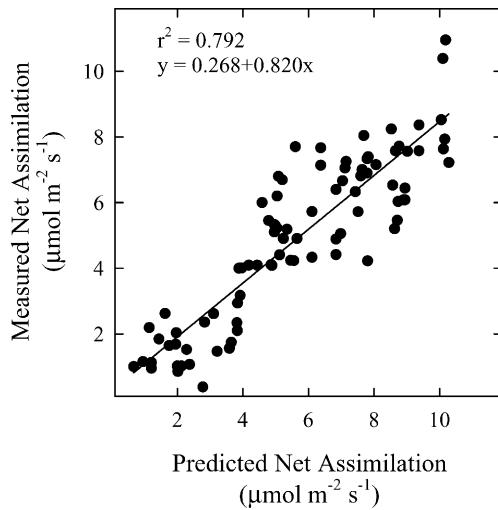


Fig. 7. Measured vs. predicted rates of net CO<sub>2</sub> assimilation ( $A_{\text{net}}$ ) for current year, 1-, and 2-year-old needles with varying levels of *P. gaemannii* infection (PD ranged from 0 to 61%) from three western Oregon Douglas-fir plantations. Each observation is the mean of six trees ( $n = 89$ ) measured in 1999.

between maximum  $A_{\text{net}}$  and *P. gaemannii* colonisation levels suggests that *P. gaemannii* can be an important regulator of  $A_{\text{net}}$  as needles age. Furthermore,

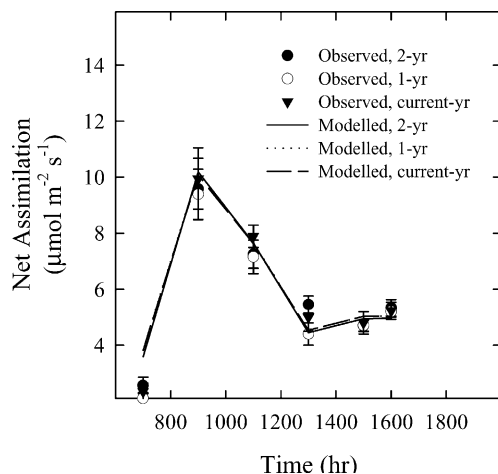


Fig. 8. Diurnal pattern of measured and predicted rates of net CO<sub>2</sub> assimilation ( $A_{\text{net}}$ ) for sprayed current year, 1- and 2-year-old needles (mean PD was 0, 0 and 4%, respectively) from the low-disease site (Mac-north) on 4 August 1998. Each observation is the mean and standard error for three trees.

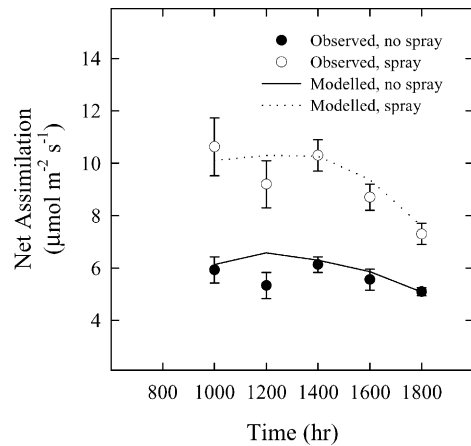


Fig. 9. Diurnal pattern of measured and predicted rates of net CO<sub>2</sub> assimilation ( $A_{\text{net}}$ ) for sprayed (mean pseudothecia density was 0%) and un-sprayed (mean PD was 15%) 1-year-old needles from the high-disease site (Beaver-south) on 7 August 1998. Each observation is the mean and standard error for three trees.

unlike other studies (e.g. Jach and Ceulemans, 2000), changes in maximum  $A_{\text{net}}$  as needles aged were not correlated with foliar nitrogen concentrations (Fig. 2).

A decline in  $A_{\text{net}}$  due to *P. gaemannii* infection was also present, with a good fit to the diurnal patterns of measured and observed values. Like  $g_s$ , diurnal rates of  $A_{\text{net}}$  were significantly reduced in diseased versus healthy trees, as shown for the high-disease site (Beaver-south) in Fig. 9.

### 2.5. Sensitivity analysis

We tested the effect of *P. gaemannii* infection on model estimates by calculating  $g_s$  and  $A_{\text{net}}$  for a typical summer day (minimum/maximum values over the 24-hr simulation period were as follows: air temperature 12.9/29.8 °C, relative humidity 60/100%, PPFD 0/1356  $\text{mmol m}^{-2} \text{s}^{-1}$ , and VPD 159/2161 Pa) over the range of observed PD and  $K_L$  values (Fig. 10). The first model run was conducted with PD levels ranging from 0 to 50% and  $K_L$  held constant at 0.11  $\text{mmol m}^{-2} \text{s}^{-1} \text{MPa}^{-1}$  (Fig. 10A); the second run was conducted with  $K_L$  values ranging from 0.13 to 0.05  $\text{mmol m}^{-2} \text{s}^{-1} \text{MPa}^{-1}$  and PD held constant at 0.0% (Fig. 10B). As shown, changes in PD had a much greater impact on gas exchange estimates. For example, over the range of PD values,  $g_s$  and  $A_{\text{net}}$  changed by 84.1 and 111.5%, respectively; and over



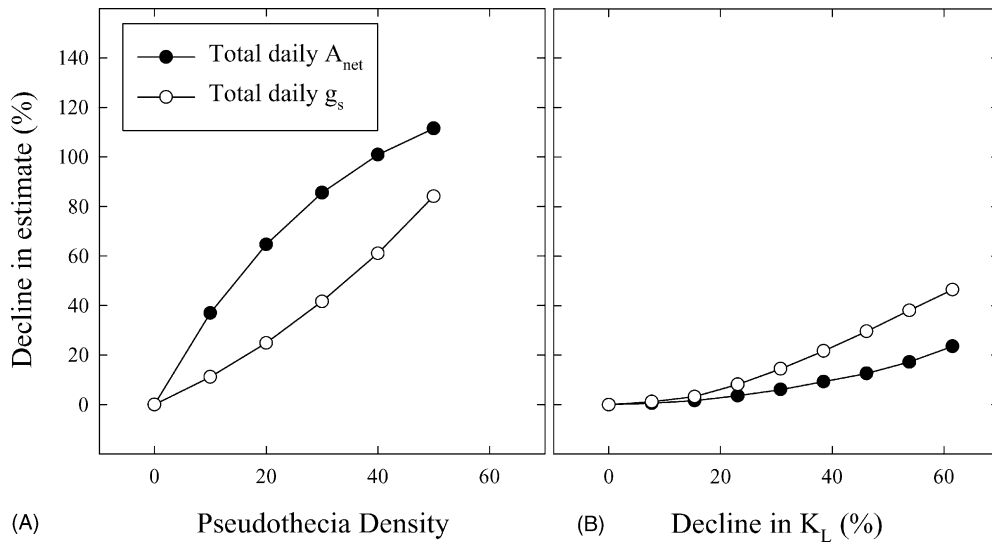


Fig. 10. Relationship between the percent decline in total daily stomatal conductance ( $g_s$ ) and net  $CO_2$  assimilation ( $A_{net}$ ) estimates and increasing PD (panel A) or declining leaf-specific hydraulic conductance ( $K_L$ ) (panel B).

the range of  $K_L$  values,  $g_s$  and  $A_{net}$  changed by only 46.5 and 23.6%, respectively.

### 3. Model simulation

As stated previously, the second objective of this study was to determine if *P. gaeumannii*'s impact on gas exchange is correlated with current growth losses in coastal Oregon Douglas-fir plantations. To determine this, branch-level estimates of  $A_{net}$  were scaled up over time and space and compared against observed volume growth for the same time period.

#### 3.1. Study sites

Long-term estimates of whole-canopy  $A_{net}$  were calculated using the model for the sprayed and un-sprayed trees on each of the north and south plots from the three Douglas-fir plantations as follows. For each hour over a 10-month sample period (1 July 1998–30 April 1999), estimates of  $g_s$  and  $A_{net}$  were calculated for needles from each age class using plot-specific values of climate data (temperature, RH, PPFD, VPD, data not shown), and needle age class-specific values of PD (Table 2), and  $K_L$  (Table 3).  $A_{net}$  estimates were integrated over time by

multiplying the sum of each hourly measurement for the 10-month period by  $3600 \text{ s h}^{-1}$ . Estimates were also scaled-up to a canopy estimate by multiplying by the whole-tree leaf area ( $\text{m}^2$ ).

#### 3.2. Whole-canopy scaling

Various techniques have been proposed to estimate whole-tree leaf areas (McDowell et al., 2002). However, in this case the best method should be easily parameterised, account for temporal changes in leaf area caused by *P. gaeumannii*-induced needle abscission, and be able to predict the relative leaf areas of all needle age classes present. Therefore, an allometric equation relating cross-sectional sapwood area ( $\text{cm}^2$ ) and leaf area ( $\text{m}^2$ ), for each of the youngest four needle age classes, was determined using healthy trees with full needle complements (i.e. 100% needle retention), and modifying these estimates in modelled trees (i.e. varying levels of *P. gaeumannii*-induced needle abscission) using visual estimates of needle retention (0–1) as follows.

One 5-year-old branch from each of the 12 healthy Mac forest trees was harvested in September 2000 for leaf area and branch diameter determinations. The leaf area for each age class ( $LA_x$ , where  $x$  denotes age class) was determined by multiplying the total

Table 2  
PD (%) and needle retention (NR, %) values for each needle age class input into model

Site code <sup>a</sup>	Date <sup>b</sup>	PD				NR			
		3-year-old	2-year-old	1-year-old	Current year	3-year-old	2-year-old	1-year-old	Current year
BNNS	7-11	–	63.0	50.7	0.0	0.0	30.1	91.2	100
BNNS	12-4	–	76.6	64.7	4.8	0.0	28.6	91.0	100
BNS	7-11	–	41.3	29.2	0.0	0.0	53.4	77.1	100
BNS	12-4	–	53.8	33.2	0.1	0.0	51.1	76.8	100
BSNS	7-11	–	–	54.5	0.0	0.0	0.0	60.5	100
BSNS	12-4	–	–	66.4	18.7	0.0	0.0	60.5	100
BSS	7-11	–	–	24.2	0.0	0.0	0.0	77.1	100
BSS	12-4	–	–	47.5	0.1	0.0	0.0	76.8	100
HNNS	7-11	–	12.2	3.6	0.0	0.0	65.8	100	100
HNNS	12-4	–	16.8	17.0	2.7	0.0	64.2	98.2	100
HNS	7-11	–	4.5	4.0	0.0	0.0	84.2	100	100
HNS	12-4	–	20.4	6.8	0.3	0.0	84	99.5	100
HSNS	7-11	–	21.3	7.2	0.0	0.0	24.2	83.3	100
HSNS	12-4	–	55.3	29.5	0.0	0.0	22.2	82.1	100
HSS	7-11	–	19.0	4.7	0.0	0.0	46.1	85.7	100
HSS	12-4	–	35.8	15.0	0.5	0.0	43.1	86	100
MNNS	7-11	2.0	1.7	2.8	0.0	100	100	100	100
MNNS	12-4	10.7	10.7	27.7	0.6	100	100	100	100
MNS	7-11	1.0	0.9	6.4	0.0	100	100	100	100
MNS	12-4	4.5	4.5	9.7	0.4	100	100	100	100
MSNS	7-11	0.5	0.2	2.7	0.0	100	100	100	100
MSNS	12-4	1.3	1.3	16.5	1.0	100	100	100	100
MSS	7-11	0.9	0.9	3.8	0.0	100	100	100	100
MSS	12-4	2.2	2.2	3.4	0.0	100	100	100	100

<sup>a</sup> The first letter corresponds to the site (B, Beaver, H, Hebo, M, Mac), the second letter is the slope (N, north, S, south), and the third and fourth letters are the spray treatment (NS, no spray, S, spray).

<sup>b</sup> Date is the month(s) of the year modelled using the listed values (7-11: July 1998–November 1998, 12-4: December 1998–April 1999).

Table 3  
Values of the percent of latewood present in sapwood (LW, %) input into model

Site code <sup>a</sup>	LW <sup>b</sup>
BNNS	63
BNS	42
BSNS	62
BSS	36
HNNS	32
HNS	25
HSNS	33
HSS	28
MNNS	35
MNS	38
MSNS	38
MSS	31

<sup>a</sup> The first letter corresponds to the site (B, Beaver, H, Hebo, M, Mac), the second letter is the slope (N, north, S, south), and the third and fourth letters are the spray treatment (NS, no spray, S, spray).

<sup>b</sup> Reported values for each site code were used for all needle age classes and months modelled.

leaf dry weight present on the branch by a projected leaf area to dry weight ratio determined from a random sub-sample of 100 needles from each branch. These branch-level, leaf area determinations were used to create an allometric equation relating branch cross-sectional area (SA, at 5 cm from junction of main stem) and leaf area. The relationship between leaf area (LA<sub>0</sub>, current year; LA<sub>1</sub>, 1-year-old, etc.) and branch cross-sectional area for the four age classes are as follows:

$$LA_0 = 0.1965 \times SA, \quad r^2 = 0.8614 \quad (17)$$

$$LA_1 = 0.1497 \times SA, \quad r^2 = 0.9039 \quad (18)$$

$$LA_2 = 0.0528 \times SA, \quad r^2 = 0.8257 \quad (19)$$

$$LA_3 = 0.0055 \times SA, \quad r^2 = 0.2186 \quad (20)$$

A whole-tree estimate of leaf area for each age class (TLA<sub>x</sub>) was calculated by measuring diameters of all living branches on each tree and summing the

estimates of  $LA_x$ . The whole-tree leaf area (TLA) is the sum of  $TLA_x$  for all needle age classes. Similar to other studies (McDowell et al., 2002), a consistent linear relationship between stem diameter at breast height (DBH) and estimates of TLA was observed:

$$TLA = 0.4045 \times DBH, \quad r^2 = 0.8983 \quad (21)$$

Finally, the relative distribution of the needle age classes for the whole-tree ( $RLA_x = TLA_x/TLA$ )

was determined to be  $47.1 \pm 1.5\%$ ,  $36.4 \pm 1.4\%$ ,  $14.2 \pm 0.9\%$ , and  $2.3 \pm 0.4\%$  for the current year, 1-, 2-, and 3-year-old needles, respectively. From the above calculations, whole-tree  $TLA_x$  for all modelled trees was estimated by Eqs. (21) and (22):

$$TLA_x = TLA \times RLA_x \times NR_x, \quad (22)$$

which could be estimated at any point in time from only DBH and needle retention ( $NR_x$ ) measurements. Estimates of leaf areas derived from Eq. (22) were

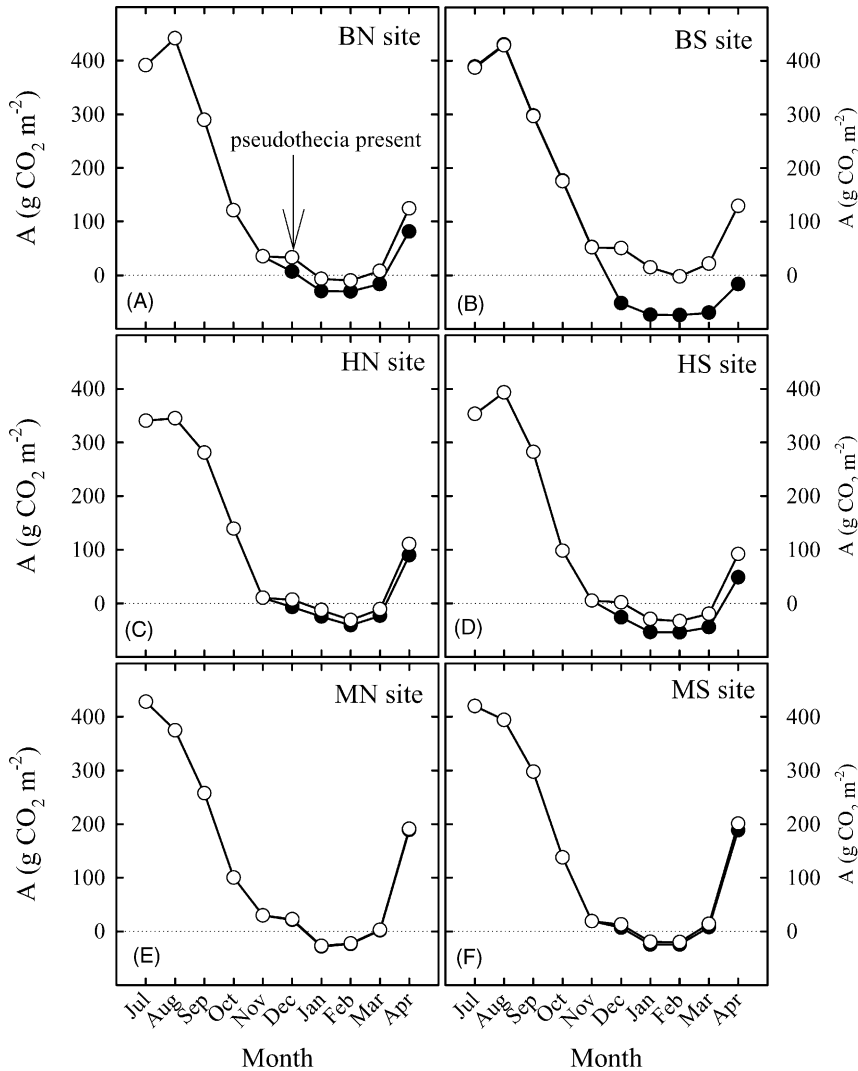


Fig. 11. Modelled total monthly value of net CO<sub>2</sub> assimilation ( $A_{net}$ ) per unit leaf area for the no spray or diseased (closed symbols) and sprayed or healthy (open symbols) current year needles. The first letter of the site code is the site (B, Beaver, H, Hebo, M, Mac), and the second letter is the slope (N, north, S, south).

verified by determining estimates of  $TLA_x$ , using the same methods outlined for the Mac trees above, for three diseased trees from the Beaver site. All estimates of  $TLA_x$  computed by the two methods were within 5% of each other (data not shown).

### 3.3. Volume growth data

An index of tree volume growth was calculated as the product of the following height and area growth measurements. In May 2000, height growth in all modelled trees was estimated by measuring the length of the 1999 internode on the main stem. Diameter growth was estimated for the same year by calculating the area of the sapwood ring, whose average radius was determined from two increment cores (north and west aspect) taken at 1.4 m above the ground.

## 4. Results

The pattern of gas exchange for current year needles over time is shown in Fig. 11.  $A_{net}$  was highest in the summer (reaching values of ca.  $400 \text{ g CO}_2 \text{ m}^{-2} \text{ LA}$ ) and declined in the winter to near or below zero. After November, when pseudothecia emerged from needle stomata,  $A_{net}$  in the sprayed and un-sprayed

trees diverged, and at the site with the highest levels of *P. gaeumannii* colonisation (Beaver-south plot) April 1999 values of  $A_{net}$  were ca.  $150$  and  $0 \text{ g CO}_2 \text{ m}^{-2} \text{ LA}$  for the sprayed and un-sprayed needles, respectively.

Estimates of the long-term carbon budgets in needles with varying levels of *P. gaeumannii* show that once infection levels were high enough, needles became a sink for carbon. Fig. 12A shows  $A_{net}$  estimates for each age class of needles from all six plots plotted against the amount of *P. gaeumannii* colonisation (pseudothecia density). Despite differences in climate, the relationship is highly significant over the 10-month simulation period, and once pseudothecia density increases above ca. 25%, carbon budgets become negative. Fig. 12B shows the relationship between  $A_{net}$  and observed needle retentions the following spring (April 2000). Based on this plot, needle retention decreased linearly as  $A_{net}$  decreased, and all needle age classes that were carbon sinks had been completely abscised by the spring of 2000.

When scaled to the canopy level all modelled trees were estimated to have positive carbon budgets (Table 4). Current year needles were the largest contributors to the carbon gain of all trees, and at the high-disease plots (Beaver) they were the only needle age class with positive carbon budgets. The relationship between the estimated whole-canopy  $A_{net}$  and

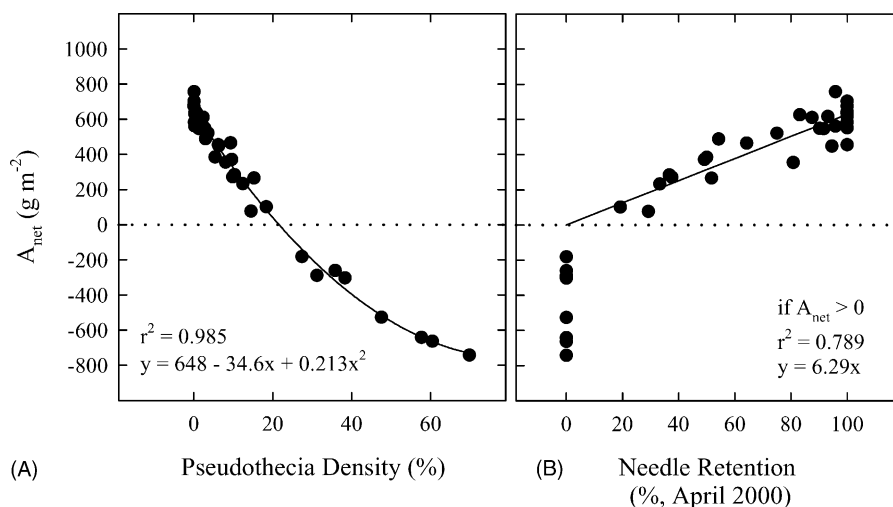


Fig. 12. Total long-term estimate of net  $\text{CO}_2$  assimilation ( $A_{net}$ ) per unit leaf area in relationship to [panel A] *P. gaeumannii* pseudothecia density (%) or [panel B] needle retention (%), April 2000).  $A_{net}$  is the modelled value for each needle age class present within a plot; pseudothecia density is the average percentage of stomata with visible pseudothecia from six trees.

Table 4

Total net CO<sub>2</sub> assimilation ( $A_{net}$ , kg) for each needle age class over the entire 10-month simulation

Age	BNNS <sup>a</sup>	BNS	BSNS	BSS
3-year-old	–	–	–	–
2-year-old	–2.7	–2.6	–	–
1-year-old	–14.7	–9.2	–6.9	–4.3
Current year	21.5	33.2	12.1	27.9
Total	4.2	21.3	5.2	23.7
	HNNS	HNS	HSNS	HSS
3-year-old	–	–	–	–
2-year-old	0.7	2.7	–1.2	–0.9
1-year-old	8.4	14.0	1.9	6.1
Current year	21.3	28.0	14.9	20.6
Total	30.4	44.7	15.6	26.2
	MNNS	MNS	MSNS	MSS
3-year-old	0.4	0.6	0.6	0.7
2-year-old	4.2	5.7	6.1	6.6
1-year-old	7.1	10.2	10.3	15.8
Current year	21.4	24.0	22.9	27.0
Total	33.1	40.5	39.9	50.1

<sup>a</sup> The first letter corresponds to the site (B, Beaver, H, Hebo, M, Mac), the second letter is the slope (N, north, S, south), and the third and fourth letters are the spray treatment (NS, no spray, S, spray). Missing values are due to needle abscission. Corresponding *P. gaemannii* infection levels are in Table 2.

volume growth rates was assessed by regression analysis (Fig. 13). Across all plots, a significant linear relationship was found ( $r^2 = 0.683$ ).

Fig. 14 shows the results of different model runs to determine the relative influence of reduced gas exchange rates per unit LA or reduced needle retention (i.e. needle abscission) on whole-canopy  $A_{net}$ . For each site, four different estimates of whole-canopy  $A_{net}$  were calculated over the 10-month simulation. These included: (i) the maximum possible  $A_{net}$  or when needle retention (NR) was set to 100% and pseudothecia density (PD) was set to 0%, (ii) the impact of needle retention, i.e. actual NR values (Table 2) with PD set to 0%, (iii) the impact of PD, i.e. actual PD values (Table 2) with NR set to 100%, and (iv) the impact of NR and PD, i.e. actual PD and NR values (Table 2). The values reported in Fig. 14 are the percent decline in the three latter simulations (ii–iv) from the maximum (i). Overall, the impact of *P. gaemannii* on needle gas exchange rates had a large influence on

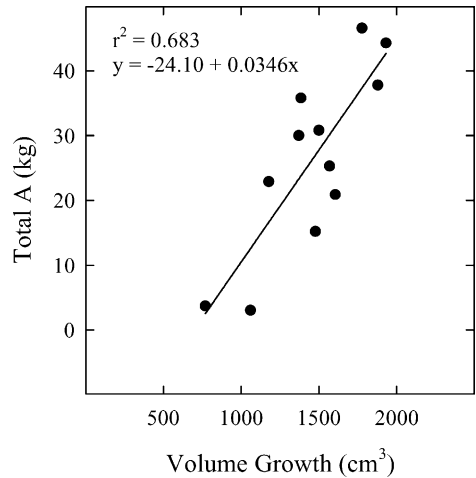


Fig. 13. Total long-term estimate of net CO<sub>2</sub> assimilation ( $A_{net}$ ) vs. volume growth. Growth parameters are the mean of six disease or six un-diseased trees from each site.

whole-canopy  $A_{net}$ . For example, at the Beaver-south plot, *P. gaemannii*'s impact on gas exchange rates reduced  $A_{net}$  by 110.6%; whereas, needle abscission and the loss of leaf area, assumed to be disease-free,

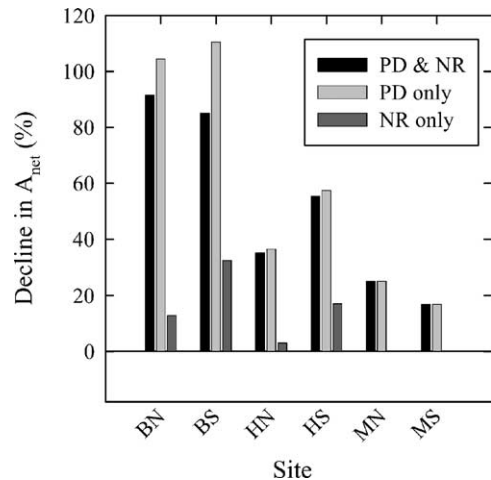


Fig. 14. Percent change in the estimate of long-term net CO<sub>2</sub> assimilation ( $A_{net}$ ) due to various disease components as compared to a model run with 100% needle retention (NR) and 0% PD. Bars are as follows: NR only compares actual and 100% NR both at 0% PD, PD only compares actual and 0% PD both at 100% NR and NR and PD compares actual PD and NR with 0% PD and 100% NR. The first letter of the site code is the site (B, Beaver, H, Hebo, M, Mac), and the second letter is the plot-aspect (N, north, S, south).

reduced  $A_{\text{net}}$  by only 32.6%. In reality, however, because the abscised needles had high levels of *P. gaemannii* infection and negative carbon budgets (i.e. not disease-free as assumed in simulation ii), their abscission mitigated the decline in whole-canopy  $A_{\text{net}}$  such that the effect of *P. gaemannii* on whole-canopy  $A_{\text{net}}$  declined to 85.1%.

## 5. Discussion

The colonisation of Douglas-fir needles by *P. gaemannii* begins in early summer when newly emerging needles are infected by deposited ascospores (Hansen and Lewis, 1997). Colonisation of needles increases over time with the emergence of pseudothecia from stomata 1–4 years later (Hansen et al., 2000). Gaseous diffusion of  $\text{CO}_2$  and  $\text{H}_2\text{O}$  is physically impaired by these fungal structures (Manter et al., 2000) and beginning in December, when pseudothecia emerged from stomata,  $A_{\text{net}}$  rates in diseased foliage are significantly reduced compared to healthy foliage.

It has been hypothesised that needle abscission begins when needles switch from being a carbon source to a carbon sink (McMurtrie et al., 1986; Cannell and Morgan, 1990). In a healthy plant, the foliar carbon budget is largely attributed to local photosynthetic rates, since mature needles import little photosynthate and respiration rates are low relative to assimilation (Sprugel et al., 1991), and nutrient content (Balster and Marshall, 2000). Two primary factors have been hypothesised to account for the decline in  $A_{\text{net}}$  of older needles: reduced light (Schoettle and Smith, 1991) and nutrient (Balster and Marshall, 2000) content. Our data demonstrate a third cause, fungal-mediated occlusion of stomata and declines in gas exchange. From our estimated carbon budgets, once approximately 25% of needle stomata contain visible *P. gaemannii* pseudothecia needles become carbon sinks. Although these needles can still actively photosynthesize during daylight hours (Manter et al., 2000), their respiration costs result in a net negative carbon balance for the year. Previous studies at the same sites used in this modelling exercise have shown that needle abscission first begins when the 25% needle colonisation level is reached (D.K. Manter, unpublished data), suggesting that needle longevity is related to carbon budgets; and a later survey (April 2000) showed that all un-

productive needle age classes had been completely abscised.

All of the carbon required for plant growth is acquired through photosynthesis; therefore, if whole-canopy  $A_{\text{net}}$  and leaf area can be measured then the amount of carbon available for growth can be estimated (e.g. Ledig, 1969). We assessed the relationship between volume growth and long-term estimates of whole-canopy  $A_{\text{net}}$ . Despite the presence of *P. gaemannii*, canopy carbon budgets of all modelled trees remained positive due to the large contribution of current year needles to the total canopy carbon budget. When *P. gaemannii* colonisation was limited by fungicide applications, carbon budgets and volume growth increased; and at all plots, a single positive linear relationship between modelled whole-canopy  $A_{\text{net}}$  and volume growth was found.

At the landscape level, fungal leaf colonists such as *P. gaemannii* may have a profound effect on net  $\text{CO}_2$  assimilation of forest canopies. For example, based on our estimates, the reported 23–50% decline in volume growth of *P. gaemannii*-infected coastal Oregon Douglas-fir trees (Maguire et al., 2002) would be equivalent to a reduction in whole-canopy  $A_{\text{net}}$  of 15.9–34.6  $\text{kg CO}_2 \text{ tree}^{-1} \text{ year}^{-1}$  (as compared to a healthy tree with an annual volume growth of 2000  $\text{cm}^3$ ). For the entire 119,500 ha of coastal Oregon forest plantations exhibiting SNC symptoms, total  $A_{\text{net}}$  would be reduced by ca. 0.5–1 million metric tons (assuming 250 trees  $\text{ha}^{-1}$ ). Furthermore, this massive reduction in carbon fixation is: (i) accomplished by a fungus whose biomass constitutes less than 2.5% of the total foliar biomass in a heavily infected tree (0.11 kg DW of *P. gaemannii* and 4.7 kg DW of LA per tree at the Beaver-south site; D.K. Manter, unpublished data), and (ii) equivalent to ca. 7.7–15.4% of the 6.5 million metric tons of  $\text{CO}_2$  emitted from Oregon's power plants in 1999 (US Department of Energy, 1999).

This paper documents the limitations on whole-canopy  $A_{\text{net}}$ , and its significant relationship to growth, imposed by the native Douglas-fir pathogen, *P. gaemannii*, in Oregon forests. The cause of the recent intensification of SNC in the PNW is unknown; however, *P. gaemannii* is widespread in both natural and managed stands of Douglas-fir. Because of Douglas-fir's high wood quality and productivity, approximately 1 million ha of plantations are currently in production



world-wide, in addition to the 14.3 million ha in the United States (Hermann and Lavender, 1999). Normally, the latent period of *P. gaemannii* lasts several years, and fungal fruiting occurs primarily on the 3-year-old and older needles. Under these conditions, *P. gaemannii* would remain a minor factor in limiting Douglas-fir forest productivity since ca. 85% of whole-canopy  $A_{\text{net}}$  occurs in the current and 1-year-old needles. Alternatively, in regions where faster *P. gaemannii* colonisation rates occur (current year and 1-year-old needles with greater than 25% of stomata with visible *P. gaemannii* pseudothecia) selection of Douglas-fir genotypes with shorter needle longevities should increase forest productivity by removal of the diseased needles that are a net drain on whole-canopy  $A_{\text{net}}$  and growth. However, since the overwhelming impact of *P. gaemannii* is on gas exchange, as long as, *P. gaemannii* is present and reaches sexual maturity (i.e. produces pseudothecia), significant declines in whole-canopy  $A_{\text{net}}$  and growth will continue.

## Acknowledgements

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